

# HIV-2 Infection and Pregnancy

Updated: January 31, 2024

Reviewed: January 31, 2024

## Panel's Recommendations

- HIV-2 infection should be considered in pregnant people who are from—or who have partners who are from—countries in which the virus is endemic and who have positive results on an HIV-1/HIV-2 antibody or HIV-1/HIV-2 antigen/antibody immunoassay. They should be tested with a supplemental HIV-1/HIV-2 antibody differentiation assay. If they have only HIV-2 infection, the test will be negative for HIV-1 antibodies and positive for HIV-2 antibodies (AII).
- Pregnant people with HIV-2 infection should be treated based on the guidelines for HIV-1 infection but using antiretroviral (ARV) drugs that are active against HIV-2. Non-nucleoside reverse transcriptase inhibitors, enfuvirtide, and fostemsavir are not active against HIV-2 and should not be used (AIII).
- No randomized clinical trials have been performed to address when to start treatment or what the optimal treatment is for HIV-2 infection (AIII). A regimen with two nucleoside reverse transcriptase inhibitors (NRTIs) and integrase strand transfer inhibitors or certain boosted protease inhibitors is recommended for all pregnant people with HIV-2 infection (AIII).
- Dolutegravir, bictegravir, raltegravir, or darunavir/ritonavir plus a dual-NRTI backbone of abacavir plus lamivudine (3TC), or tenofovir disoproxil fumarate or tenofovir alafenamide plus emtricitabine or 3TC, are recommended for treating HIV-2 - infection alone in pregnant people and in people who are trying to conceive (AIII). Zidovudine (ZDV) plus 3TC can be used as an alternative dual-NRTI backbone. See [Recommendations for Use of Antiretroviral Drugs During Pregnancy](#) and [Appendix C: Antiretroviral Counseling Guide for Health Care Providers](#).
- If a pregnant individual is already receiving antiretroviral therapy with drugs that are active against HIV-2, treatment should be continued (AIII).
- As with HIV-1, the possibility of hepatitis B virus/HIV-2 coinfection should be considered when choosing an ARV regimen to treat HIV-2 (AI) (see [Hepatitis B Virus/HIV Coinfection](#)).
- All infants born to people with HIV-2 infection (without HIV-1 infection) should receive a 4-week ZDV prophylactic regimen (BIII) (see [Table 10. Neonatal Antiretroviral Management According to Risk of HIV Infection in the Newborn](#) and [Table 11. Antiretroviral Drug Dosing Recommendations for Newborns](#)).
- People with HIV-2 infection should receive patient-centered, evidence-based counseling to support shared decision-making about infant feeding options prior to and during pregnancy; counseling and plans for infant feeding should be reviewed again after delivery (AIII) (see [Infant Feeding for Individuals with HIV in the United States](#)).

**Rating of Recommendations:** A = Strong; B = Moderate; C = Optional

**Rating of Evidence:** I = One or more randomized trials with clinical outcomes and/or validated laboratory endpoints;

II = One or more well-designed, nonrandomized trials or observational cohort studies with long-term clinical outcomes;

III = Expert opinion

HIV-2 infection is endemic in West African countries, including Burkina Faso, Cape Verde, The Gambia, Ghana, Guinea, Guinea-Bissau, Ivory Coast, Liberia, Mali, Mauritania, Niger, Nigeria, Sao Tome, Senegal, Sierra Leone, and Togo. It is also endemic in Angola, Mozambique, and parts of India.<sup>1-4</sup> It also occurs in countries that have large numbers of immigrants from these regions, such as France and Portugal.<sup>5</sup>

HIV-2 remains rare in the United States. According to the National HIV Surveillance System, 327,700 diagnoses of HIV were recorded in the United States from 2010 to 2017, of which 198 (0.06%) met the criteria for HIV-2 (HIV-2 infection alone, n = 102; dual HIV-1 and HIV-2, n = 11; probable but unconfirmed HIV-2, n = 85).<sup>6</sup> Among these cases, 99 women had diagnoses of

confirmed or probable HIV-2, and 9 of these women had evidence of pregnancy at or after their diagnosis. No perinatal HIV-2 transmissions were reported. HIV-2 infection should be suspected in pregnant people who are from—or who have partners from—countries in which the disease is endemic and who have positive results on an HIV-1/HIV-2 antibody or HIV-1/HIV-2 antigen/antibody immunoassay. They should be tested with a supplemental HIV-1/HIV-2 antibody differentiation immunoassay. If they have only HIV-2 infection, the test will be negative for HIV-1 antibodies and positive for HIV-2 antibodies. In rare instances, a person may have dual infection with HIV-1 and HIV-2, and both tests will be positive.

In 2014, the Centers for Disease Control and Prevention (CDC) released a new HIV testing algorithm. The first step in this algorithm is performing an HIV-1/HIV-2 antigen/antibody combination assay on serum or plasma (e.g., Abbott Architect HIV Ag/Ab combo assay, Bio-Rad GS Combo Ag/Ab EIA, Alere Determine).<sup>7</sup> This test does not distinguish between HIV-1 antibodies and HIV-2 antibodies. Specimens that are reactive on this test must be tested with a U.S. Food and Drug Administration (FDA)—approved antibody assay to distinguish HIV-1 antibodies from HIV-2 antibodies. The FDA-approved HIV-2 antibody supplemental test Geenius (Bio-Rad Laboratories) is used as part of the CDC-recommended HIV laboratory testing algorithm.

Viral load assays for HIV-2 are not commercially available, but they may be available under research protocols. The [University of Washington](#)<sup>8</sup> and the [New York State Department of Health, Wadsworth Center](#)<sup>9</sup> also offer HIV-2 viral load assays. The University of Washington accepts specimens forwarded from laboratories, such as Quest Diagnostics. All HIV-2 cases should be reported to the HIV surveillance program of the state or local health department, which can arrange for additional confirmatory testing for HIV-2 by the CDC.<sup>10</sup> No validated HIV-2 genotype or phenotype resistance assays are available in the United States. HIV-2 genotypic resistance assays are available for research use only at the University of Washington. Drug resistance against HIV-2 can be determined using the [HIV-2 EU resistance tool](#), and another [French resistance tool](#).<sup>11,12</sup>

HIV-2 has a longer asymptomatic phase than HIV-1, with a slower progression to AIDS. However, without effective antiretroviral therapy (ART), HIV-2 will progress to AIDS and death in the majority of individuals over time.<sup>13</sup> The most common mode of HIV-2 transmission is through sex. HIV-2 is less infectious than HIV-1, with a fivefold lower rate of sexual transmission and 20-fold to 30-fold lower rate of perinatal transmission.<sup>3,14,15</sup> Several studies confirm that rates of perinatal transmission of HIV-2 are low with and without interventions (0% to 4%), which may be a result of reduced plasma viral loads, higher CD4 T lymphocyte (CD4) cell counts, and less cervical viral shedding in women with HIV-2 infection than in women with HIV-1 infection.<sup>16-19</sup> In a meta-analysis comparing perinatal transmission of HIV-1 and HIV-2, the pooled incidence of HIV-2 perinatal transmission was 0.2% (95% confidence interval [CI], 0.03% to 1.47%) among antiretroviral-naïve pregnant women.<sup>20</sup>

HIV-2 also can be transmitted through breastfeeding. HIV-2 infection does not protect against HIV-1, and dual infection—which carries the same prognosis as HIV-1 infection alone—can occur.<sup>21</sup>

## Recommended Antiretroviral Therapy for Pregnant People with HIV-2 Infection

Pregnant people with HIV-2 infection should be treated according to the guidelines for patients with HIV-1 infection alone, although clinicians should make sure that the chosen antiretroviral (ARV) regimen is also appropriate for treatment of HIV-2. Once treatment is started, ART should be continued postpartum as is recommended for all patients with HIV-1. A systematic review analyzed

data collected from 1996 to 2012 on treatment outcomes among nonpregnant patients with HIV-2. The review reported a heterogeneity of treatment outcomes among patients who initiated ART, especially in resource-limited settings.<sup>22</sup> Non-nucleoside reverse transcriptase inhibitors (NNRTIs), enfuvirtide, and fostemsavir are not active against HIV-2 and **should not be used** for treatment or prophylaxis.<sup>23,24</sup> The integrase strand transfer inhibitors (INSTIs) raltegravir (RAL), elvitegravir, dolutegravir (DTG), bictegravir (BIC), and cabotegravir (CAB) are effective against HIV-2.<sup>25,26</sup> Although DTG may be able to rescue a failing RAL-based regimen in a person with HIV-2 infection, a study has reported the emergence of DTG-resistance mutations in people with HIV-2 infection.<sup>27</sup> The CCR5 antagonist maraviroc appears to be active against some strains of HIV-2, although no approved assays exist to determine HIV-2 co-receptor tropism.<sup>28,29</sup> HIV-2 drug resistance has been documented with various ARV drugs.<sup>30,31</sup> Among 47 ART-naive people with HIV-2, ultradeep sequencing showed that three people displayed plasma viruses with a resistance-associated mutation (RAM) above the 20% detection threshold, with a prevalence of transmitted drug resistance for nucleoside reverse transcriptase inhibitors (NRTIs) of 7.9% (95% CI, 0.0% to 16.5%). No RAM above the 20% detection threshold was found for protease inhibitors (PIs) or INSTIs.<sup>32</sup> For patients with multidrug-resistant virus, ibalizumab and lenacapavir demonstrate *in vitro* potency against HIV-2 and may be considered; these drugs are not recommended except in special circumstances for use in pregnancy.

HIV-2 has variable susceptibility to PIs, with lopinavir (LPV) and darunavir (DRV) having the most activity.<sup>33</sup>

The care of pregnant people with HIV-2 infection alone has been based on expert opinion. A regimen with two NRTIs and an INSTI or a ritonavir-boosted PI currently is recommended for all pregnant people with HIV-2 infection. The following regimens can be used to treat HIV-2, based on the available efficacy and safety data on these drugs from clinical trials of pregnant people with HIV-1 infection:

- DTG, BIC, RAL, or darunavir/ritonavir plus a dual-NRTI backbone of abacavir plus lamivudine (3TC), or tenofovir disoproxil fumarate or tenofovir alafenamide plus emtricitabine or 3TC are the recommended regimens for treating HIV-2 infection alone in pregnant people and people who are trying to conceive. See [Recommendations for the Use of Antiretroviral Drugs During Pregnancy: Overview](#), [Table 6. What to Start: Initial Antiretroviral Regimens During Pregnancy for People Who Are Antiretroviral-Naive](#), [Table 7. Situation-Specific Recommendations for Use of Antiretroviral Drugs in Pregnant People and Nonpregnant People Who Are Trying to Conceive](#), and [Appendix C: Antiretroviral Counseling Guide for Health Care Providers](#).<sup>34</sup>
- Zidovudine (ZDV) plus 3TC can be used as an alternative dual-NRTI backbone.
- NNRTIs **should not be used** because they are not active against HIV-2.

If a pregnant individual is already receiving ART with drugs that are active against HIV-2, treatment should be continued.

When monitoring the plasma viral loads and CD4 counts in pregnant people with HIV-2 infection, clinicians should follow the guidelines outlined for people with HIV-1 infection (see [Initial Evaluation and Continued Monitoring of HIV-Related Assessments During Pregnancy](#)). However, disease progression can occur in the setting of undetectable HIV-2 plasma viral load. Patients who have HIV-2 plasma viral loads that are below the limits of detection should still have routine CD4

counts and clinical monitoring (see [Plasma HIV-1 RNA \[Viral Load\] and CD4 Count Monitoring](#) in the [Adult and Adolescent Antiretroviral Guidelines](#)).

No data are available to address whether treatment should be continued after pregnancy in people with HIV-2 infection alone. To date, no randomized trials have addressed the question of an optimal treatment strategy for HIV-2 infection. The [Adult and Adolescent Antiretroviral Guidelines](#) recommend that all patients with HIV-2 infection should be treated using the guidelines provided for patients with HIV-1 infection, see [HIV-2 Infection](#).

All infants born to people with HIV-2 who do not have HIV-1 should receive a 4-week ZDV prophylaxis regimen (see [Table 10. Neonatal Antiretroviral Management According to Risk of HIV Infection in the Newborn](#) and [Table 11. Antiretroviral Drug Dosing Recommendations for Newborns](#)). The possible risks and benefits of ARV prophylaxis should be discussed with the [birthing parents](#). As noted above, rates of perinatal transmission of HIV-2 are low with and without interventions, and it is unclear whether infants born to people with undetectable HIV-2 viral loads will benefit from ARV prophylaxis. However, monitoring HIV-2 plasma viral loads [in birthing parents](#) and receiving the results in a timely manner can be difficult because plasma samples must be sent to the University of Washington or the New York State Department of Health. Therefore, the Panel on Treatment of HIV During Pregnancy and Interventions to Prevent Perinatal HIV Transmission recommends that all infants born to [pregnant people](#) with HIV-2 receive prophylaxis. ZDV prophylaxis is recommended in this clinical situation because nevirapine lacks activity against HIV-2. Guidance on ARV prophylaxis for infants born to individuals with HIV-1 and HIV-2 infection is available in [Table 10. Neonatal Antiretroviral Management According to Risk of HIV Infection in the Newborn](#) and [Table 11. Antiretroviral Drug Dosing Recommendations for Newborns](#).

No data exist on the impact of scheduled cesarean delivery on HIV-2 perinatal transmission. The risk to infants from breastfeeding is lower for HIV-2 than for HIV-1, but the risk is not zero.<sup>17</sup> Individuals with HIV-2 infection should receive patient-centered, evidence-based counseling to support shared decision-making about infant feeding. Counseling about infant feeding should begin prior to conception or as early as possible in pregnancy; information about and plans for infant feeding should be reviewed throughout pregnancy and again after delivery (see [Infant Feeding for Individuals with HIV in the United States](#)).

Infants born to mothers with HIV-2 should be tested for HIV-2 infection with HIV-2-specific virologic assays at time points similar to those used for HIV-1 testing; see [Diagnosis of HIV Infection in Infants and Children](#).<sup>35</sup> Quantitative HIV-2 plasma RNA viral load testing for clinical care is available from the [University of Washington](#)<sup>8</sup> and the [New York State Department of Health](#).<sup>9</sup> Antibody testing of infants (e.g., with the Bio-Rad Laboratories Multispot HIV-1/HIV-2 test) also can be performed at age 18 months to confirm clearance of HIV-2 antibodies.

## References

1. De Cock KM, Brun-Vezinet F. Epidemiology of HIV-2 infection. *AIDS*. 1989;3 Suppl 1:S89-95. Available at: <https://pubmed.ncbi.nlm.nih.gov/2514761>.
2. De Cock KM, Adjorlolo G, Ekpi E, et al. Epidemiology and transmission of HIV-2. Why there is no HIV-2 pandemic. *JAMA*. 1993;270(17):2083-2086. Available at: <https://pubmed.ncbi.nlm.nih.gov/8147962>.
3. Campbell-Yesufu OT, Gandhi RT. Update on human immunodeficiency virus (HIV)-2 infection. *Clin Infect Dis*. 2011;52(6):780-787. Available at: <https://pubmed.ncbi.nlm.nih.gov/21367732>.
4. Heitzinger K, Sow PS, Dia Badiane NM, et al. Trends of HIV-1, HIV-2 and dual infection in women attending outpatient clinics in Senegal, 1990-2009. *Int J STD AIDS*. 2012;23(10):710-716. Available at: <https://pubmed.ncbi.nlm.nih.gov/23104745>.
5. Cazein F, Lot F, Pillonel J, et al. HIV and AIDS surveillance in France, 2006. *Bull Epidemiol Hebd*. 2007(46-47):386-393.
6. Peruski AH, Wesolowski LG, Delaney KP, et al. Trends in HIV-2 diagnoses and use of the HIV-1/HIV-2 differentiation test—United States, 2010-2017. *MMWR Morb Mortal Wkly Rep*. 2020;69(3):63-66. Available at: <https://pubmed.ncbi.nlm.nih.gov/31971928>.
7. Centers for Disease Control and Prevention. Laboratory testing for the diagnosis of HIV iInfection: updated recommendations. 2014. Available at: <http://stacks.cdc.gov/view/cdc/23447>
8. Chang M, Gottlieb GS, Dragavon JA, et al. Validation for clinical use of a novel HIV-2 plasma RNA viral load assay using the Abbott m2000 platform. *J Clin Virol*. 2012;55(2):128-133. Available at: <https://pubmed.ncbi.nlm.nih.gov/22832059>.
9. Styer LM, Miller TT, Parker MM. Validation and clinical use of a sensitive HIV-2 viral load assay that uses a whole virus internal control. *J Clin Virol*. 2013;58 Suppl 1:e127-133. Available at: <https://pubmed.ncbi.nlm.nih.gov/24342472>.
10. Branson BM, Pandori M. 2012 HIV diagnostics conference: the molecular diagnostics perspective. *Expert Rev Mol Diagn*. 2013;13(3):243-245. Available at: <https://pubmed.ncbi.nlm.nih.gov/23570401>.
11. Charpentier C, Camacho R, Ruelle J, et al. HIV-2EU: supporting standardized HIV-2 drug resistance interpretation in Europe. *Clin Infect Dis*. 2013;56(11):1654-1658. Available at: <https://pubmed.ncbi.nlm.nih.gov/23429380>.
12. Berzow D, Descamps D, Obermeier M, et al. Human Immunodeficiency Virus-2 (HIV-2): a summary of the present standard of care and treatment options for individuals living with HIV-2 in Western Europe. *Clin Infect Dis*. 2021;72(3):503-509. Available at: <https://pubmed.ncbi.nlm.nih.gov/32227124>.

13. Esbjornsson J, Mansson F, Kvist A, et al. Long-term follow-up of HIV-2-related AIDS and mortality in Guinea-Bissau: a prospective open cohort study. *Lancet HIV*. 2018;S2352-3018(18):30254-30256. Available at: <https://pubmed.ncbi.nlm.nih.gov/30392769>.
14. Kanki PJ, Travers KU, S MB, et al. Slower heterosexual spread of HIV-2 than HIV-1. *Lancet*. 1994;343(8903):943-946. Available at: <https://pubmed.ncbi.nlm.nih.gov/7909009>.
15. Matheron S, Courpotin C, Simon F, et al. Vertical transmission of HIV-2. *Lancet*. 1990;335(8697):1103-1104. Available at: <https://pubmed.ncbi.nlm.nih.gov/1970407>.
16. O'Donovan D, Ariyoshi K, Milligan P, et al. Maternal plasma viral RNA levels determine marked differences in mother-to-child transmission rates of HIV-1 and HIV-2 in The Gambia. MRC/Gambia government/university college London medical school working group on mother-child transmission of HIV. *AIDS*. 2000;14(4):441-448. Available at: <https://pubmed.ncbi.nlm.nih.gov/10770548>.
17. Burgard M, Jasseron C, Matheron S, et al. Mother-to-child transmission of HIV-2 infection from 1986 to 2007 in the ANRS French Perinatal Cohort EPF-CO1. *Clin Infect Dis*. 2010;51(7):833-843. Available at: <https://pubmed.ncbi.nlm.nih.gov/20804413>.
18. Adjourlolo-Johnson G, De Cock KM, Ekpini E, et al. Prospective comparison of mother-to-child transmission of HIV-1 and HIV-2 in Abidjan, Ivory Coast. *JAMA*. 1994;272(6):462-466. Available at: <https://pubmed.ncbi.nlm.nih.gov/8040982>.
19. Andreasson PA, Dias F, Naucler A, et al. A prospective study of vertical transmission of HIV-2 in Bissau, Guinea-Bissau. *AIDS*. 1993;7(7):989-993. Available at: <https://pubmed.ncbi.nlm.nih.gov/8357558>.
20. Ter Schiphorst E, Hansen KC, Holm M, Honge BL. Mother-to-child HIV-2 transmission: comparison with HIV-1 and evaluation of factors influencing the rate of transmission. A systematic review. *Trans R Soc Trop Med Hyg*. 2022;116(5):399-408. Available at: <https://pubmed.ncbi.nlm.nih.gov/34791488>.
21. Prince PD, Matser A, van Tienen C, et al. Mortality rates in people dually infected with HIV-1/2 and those infected with either HIV-1 or HIV-2: a systematic review and meta-analysis. *AIDS*. 2014;28(4):549-558. Available at: <https://pubmed.ncbi.nlm.nih.gov/23921613>.
22. Ekouevi DK, Tchounga BK, Coffie PA, et al. Antiretroviral therapy response among HIV-2 infected patients: a systematic review. *BMC Infect Dis*. 2014;14:461. Available at: <https://pubmed.ncbi.nlm.nih.gov/25154616>.
23. Tuaillet E, Gueudin M, Lemee V, et al. Phenotypic susceptibility to nonnucleoside inhibitors of virion-associated reverse transcriptase from different HIV types and groups. *J Acquir Immune Defic Syndr*. 2004;37(5):1543-1549. Available at: <https://pubmed.ncbi.nlm.nih.gov/15577405>.

24. Poveda E, Rodes B, Toro C, Soriano V. Are fusion inhibitors active against all HIV variants? *AIDS Res Hum Retroviruses*. 2004;20(3):347-348. Available at: <https://pubmed.ncbi.nlm.nih.gov/15117459>.
25. Le Hingrat Q, Collin G, Le M, et al. A new mechanism of resistance of HIV-2 to integrase inhibitors: a 5 amino-acids insertion in the integrase C-terminal domain. *Clin Infect Dis*. 2018;69(4):657-667. Available at: <https://pubmed.ncbi.nlm.nih.gov/30383215>.
26. Smith RA, Raugi DN, Wu VH, et al. Comparison of the antiviral activity of bictegravir against HIV-1 and HIV-2 isolates and integrase inhibitor-resistant HIV-2 mutants. *Antimicrob Agents Chemother*. 2019;63(5):e00014-00019 Available at: <https://pubmed.ncbi.nlm.nih.gov/30803972>.
27. Requena S, Trevino A, Cabezas T, et al. Drug resistance mutations in HIV-2 patients failing raltegravir and influence on dolutegravir response. *J Antimicrob Chemother*. 2017;72(7):2083-2088. Available at: <https://pubmed.ncbi.nlm.nih.gov/28369593>.
28. Borrego P, Taveira N. HIV-2 susceptibility to entry inhibitors. *AIDS Rev*. 2013;15(1):49-61. Available at: <https://pubmed.ncbi.nlm.nih.gov/23449229>.
29. Visseaux B, Charpentier C, Hurtado-Nedelec M, et al. In vitro phenotypic susceptibility of HIV-2 clinical isolates to CCR5 inhibitors. *Antimicrob Agents Chemother*. 2012;56(1):137-139. Available at: <https://pubmed.ncbi.nlm.nih.gov/22064539>.
30. Charpentier C, Visseaux B, Benard A, et al. Transmitted drug resistance in French HIV-2-infected patients. *AIDS*. 2013;27(10):1671-1674. Available at: <https://pubmed.ncbi.nlm.nih.gov/23595155>.
31. Menendez-Arias L, Alvarez M. Antiretroviral therapy and drug resistance in human immunodeficiency virus type 2 infection. *Antiviral Res*. 2014;102:70-86. Available at: <https://pubmed.ncbi.nlm.nih.gov/24345729>.
32. Storto A, Visseaux B, Bertine M, et al. Minority resistant variants are also present in HIV-2-infected antiretroviral-naive patients. *J Antimicrob Chemother*. 2018;73(5):1173-1176. Available at: <https://pubmed.ncbi.nlm.nih.gov/29415189>.
33. Desbois D, Roquebert B, Peytavin G, et al. In vitro phenotypic susceptibility of human immunodeficiency virus type 2 clinical isolates to protease inhibitors. *Antimicrob Agents Chemother*. 2008;52(4):1545-1548. Available at: <https://pubmed.ncbi.nlm.nih.gov/18227188>.
34. Reeves I, Cromarty B, Deayton J, et al. British HIV Association guidelines for the management of HIV-2 2021. *HIV Med*. 2021;22 Suppl 4:1-29. Available at: <https://pubmed.ncbi.nlm.nih.gov/34927347>.
35. Panel on Antiretroviral Therapy and Medical Management of Children Living with HIV. Guidelines for the use of antiretroviral agents in pediatric HIV infection. 2021. Available at: <https://clinicalinfo.hiv.gov/sites/default/files/guidelines/documents/pediatric-arv/guidelines-pediatric-arv.pdf>.