

Epidemiology

Talaromycosis is an invasive fungal infection caused by the dimorphic fungus *Talaromyces marneffeii* (formerly *Penicillium marneffeii*) which is endemic in Southeast Asia (in northern Thailand, Vietnam, and Myanmar), East Asia (in southern China, Hong Kong, and Taiwan), and South Asia (in northeastern India) (see the geographic distribution of talaromycosis in Figure 1).¹⁻⁴ *Talaromyces marneffeii* was formerly classified under the *Penicillium* subgenus *Biverticillium* based on morphological characteristics. In 2011, the subgenus *Biverticillium* was found to form a monophyletic group with *Talaromyces* that is distinct from *Penicillium*, and was taxonomically unified with the *Talaromyces* genus.⁵ Hence, *Penicillium marneffeii* was changed to *Talaromyces marneffeii*, and the disease penicilliosis is now called talaromycosis.

HIV is a major risk factor for talaromycosis in highly-endemic regions, accounting for approximately 88% of disease.² The fungus is also a major cause of HIV-associated opportunistic infections in these regions, making up to 16% of hospital admissions due to AIDS,^{2,3,6-8} and is a leading cause of HIV-associated blood stream infection and death in Vietnam and southern China.^{6,9-11} Infection occurs predominantly in individuals who have very advanced HIV disease with a CD4 T lymphocyte (CD4) cell count of <100 cells/mm³.^{2,3,12} Talaromycosis is increasingly diagnosed in immunocompromised individuals who are returning travelers or immigrants from the endemic regions, and has been reported in Japan, Australia, Belgium, France, Germany, the Netherlands, Sweden, Switzerland, the United Kingdom, Oman in the Middle East, and the United States.^{13,14} Talaromycosis is increasingly recognized in individuals who have a primary immunodeficiency condition (e.g., idiopathic CD4 lymphopenia, anti-interferon-gamma autoantibody-associated immunodeficiency, conditions due to mutations in CYBB, CD40L, or gain-of-function mutation in STAT1/STAT3 pathways); secondary immunodeficiency conditions (e.g., autoimmune diseases in patients on corticosteroids and/or other immunosuppressive therapy, solid and hematological malignancies, solid organ transplantation, hematopoietic stem cell transplantation, and therapy with novel target therapies such as monoclonal antibodies against CD20 and kinase inhibitors).¹⁵ Talaromycosis-related mortality despite antifungal therapy in patients both with and without HIV is up to 30%.^{2,3,12,16,17}

Similar to other endemic mycoses, talaromycosis is a saprozoontic infection, meaning the transmissible source has a reservoir both in an abiotic environment and in an animal host. The wild bamboo rat in highland areas in the endemic regions is the known animal reservoir of *T. marneffeii*;^{18,19} however, case-control studies suggest that human infection results from inhalation of fungal spores released from a soil-related environmental reservoir (plants and farmed animals) rather than from direct bamboo rat to human transmission.^{20,21} Talaromycosis incidence increased 30% to 50% during the rainy months in southern Vietnam and northern Thailand,^{3,22} and was associated with increased humidity and not precipitation,^{23,24} suggesting that humidity facilitates an expansion of the environmental reservoir resulting in increased exposure to the fungus. Reactivation of latent infections has been demonstrated in non-autochthonous cases with a history of remote travel to the endemic countries and can occur many years after exposure.^{13,14,25} One case of presumed laboratory-acquired talaromycosis was reported in an African man with HIV while at the Pasteur Institute in Paris;²⁶ however, laboratory-acquired infection has never been reported from the endemic regions. Donor-acquired transmission has been reported in a lung-transplant recipient from Belgium.²⁷

Clinical Manifestations

Disseminated infection involving multiple organ systems is the most common manifestation of talaromycosis in patients with advanced HIV disease. The infection frequently begins as a subacute illness characterized by fever, weight loss, hepatosplenomegaly, lymphadenopathy, and respiratory and gastrointestinal abnormalities.^{3,28} These clinical features are non-specific and are indistinguishable from those of disseminated tuberculosis, other systemic mycoses, or infections due to intracellular pathogens such as *Salmonella* species.

Skin lesions are the most specific but late manifestations of talaromycosis, with central-necrotic papules appearing on the face, trunk, and extremities occurring in 40% to 70% of patients.^{1,3,29} Pulmonary involvement manifested as cough or shortness of breath occurs in 40% of patients. Gastrointestinal involvement presenting as diarrhea or abdominal pain occurs in 30% of patients. Significant hepatosplenomegaly is present in 70% of patients and together with intra-abdominal lymphadenopathy cause abdominal distention and pain.^{3,7} Meningoencephalitis is a rare manifestation, occurring in <1% of patients, and has a rapid disease course with a mortality of 80%.³⁰ Concurrent infections with other opportunistic pathogens occur in up to 60% of patients, with oropharyngeal candidiasis being the most common.²

Tuberculosis coinfection is common (occurring in up to 22% of patients in highly endemic regions) and complicates disease management because of itraconazole and rifampin drug interactions.³

Common laboratory findings associated with talaromycosis include anemia and thrombocytopenia due to bone marrow infiltration. Anemia can be profound and may require multiple red cell transfusions. Elevation of aminotransferase is common, with serum aspartate aminotransferase (AST) over alanine aminotransferase (ALT) ratio of approximately 2.³

The median CD4 count in multiple cohorts is <50 cells/mm³.^{2,3}

The chest radiographical findings are broad, ranging from diffuse interstitial disease to reticulonodular infiltrates to alveola infiltrates causing respiratory failure.³¹

Diagnosis

A diagnosis of talaromycosis should be considered in all patients with HIV with CD4 count <100 cells/mm³ who have traveled to or have lived in talaromycosis-endemic areas and present with a systemic infection involving the reticuloendothelial system (i.e., lymph nodes, liver, spleen, and bone marrow).

Skin lesions in talaromycosis have typical central-necrotic appearance and can be a diagnostic sign. However, skin lesions are a late manifestation of talaromycosis and are absent in up to 60% of patients.^{1,3,29} The current diagnostic methods for talaromycosis are still based on conventional microscopy, histology, and culture. Culture results usually return within 4 days to 5 days but can take up to 14 days. Diagnostic delay, particularly in patients presenting without fever or skin lesions, is associated with increased mortality.^{2,3,15,32} Antigen detection and polymerase chain reaction (PCR)-based methods are promising rapid diagnostics currently being evaluated.

Microscopy, Histology, and Culture are the Current Gold Standard Diagnostic Methods

A presumptive diagnosis of talaromycosis can be made based on the microscopic examination of Giemsa-, Wright-, or Gomori Methenamine Silver (GMS)-stained samples of skin lesion scrapings, lymph node aspirate, bone marrow aspirate, or tissue sections showing round to oval extracellular and intra-macrophage yeast-like organisms measuring 3 to 6 µm in diameter. Identification of a clear midline septum in a dividing yeast cell is what distinguishes *T. marneffeii* from *Histoplasma* or *Candida* species.¹ In some patients, the fungus can be identified by microscopic examination of a Wright's-stained peripheral blood smear.³³

A definitive diagnosis of talaromycosis can be made by the histopathologic demonstration of the organism in biopsy specimens. There are three histopathological forms. The granulomatous reaction is formed by histiocytes, lymphocytes, and plasma, epithelioid, and giant cells and can be seen in reticuloendothelial organs in patients who are HIV-negative or immunocompetent. The suppurative reaction develops with the joining of multiple abscesses seen in the lung and subcutaneous tissues of immunocompetent patients. The anergic and necrotizing reaction is characterized by focal necrosis surrounded by distended histiocytes containing proliferating fungi seen in the lung, liver, and spleen of immunocompromised patients.³⁴

Most frequently a definitive diagnosis of talaromycosis is based on isolation of the organism from cultures of clinical specimens.

Compared to other endemic dimorphic fungi, *T. marneffeii* grows more readily in standard BACTEC blood culture media and Sabouraud dextrose agar, but takes 5 to 14 days to grow and to demonstrate temperature dimorphism. At 25°C to 30°C, the fungus grows as a mold producing yellow-green colonies with sulcate folds and a red diffusible pigment in the media. Microscopically, filamentous hyphae with characteristic spore-bearing structures called conidiophores and conidia can be seen. At 32°C to 37°C, the fungus makes the morphological transition from a mold to a yeast, producing tan colored colonies without a red diffusible pigment. In laboratory media, only the transitional sausage-shaped cells can be seen microscopically. The round-to-oval yeast cells are only seen in natural tissue.¹

Culture yield is the highest from bone marrow (100%), followed by skin lesions (90%) and blood (70%).^{3,35} Less commonly, talaromycosis has been diagnosed from sputum, pleural fluid, peritoneal fluid, cerebrospinal fluid, pericardium fluid, stool, and urine.

Molecular Diagnosis

Molecular diagnostics for talaromycosis have been based on PCR amplification and sequence identification of specific regions within the fungal ribosome's internally transcribed spacer regions, the 5.8S rRNA, and the 18S rRNA genes of *T. marneffeii*.³⁶⁻³⁹ These assays have high specificity (100%), but limited sensitivity (60% to 70%). At present, none of the real-time PCR assays has been prospectively validated, standardized, or commercially developed for clinical use.

Antigen Detection

The commercial assay for the detection of *Aspergillus* galactomannan cross reacts with *T. marneffeii* and has a sensitivity of 95.8% (23 of 24 patients with culture-positive talaromycosis were correctly identified) and a specificity of 90.9% (30 of 33 people without talaromycosis were correctly identified) for the detection of talaromycosis (at cutoff index = 1.0).⁴⁰ However, the galactomannan test also cross reacts with other endemic fungi such as *Histoplasma* and *Blastomyces* and has not been evaluated prospectively.

The Mp1p ELISA has been shown to be more sensitive than blood culture (in 372 culture-proven talaromycosis cases, sensitivity was 86.3% for the Mp1p ELISA and 74% for blood culture) and is highly specific (98.1% specificity in 338 healthy controls and 179 patients without HIV, but with other infections).⁴¹ This assay was used to screen a large serum bank of 8,131 patients with HIV in Guangzhou, China and showed a Mp1p antigenemia prevalence of 9.4%, with prevalence of antigenemia increased from 4.5% to 28.4% as the CD4 count decreased from 200 cells/mm³ to 50 cells/mm³, demonstrating a significant burden of disease in southern China.²⁴ In Vietnam, the Mp1p ELISA identified 4.2% antigenemia in 1,123 asymptomatic patients initiating antiretroviral therapy (ART) in 22 HIV clinics across Vietnam who had a CD4 count <100 cells/mm³. Antigenemia was found to be independently associated with 12-month mortality.⁴² These data demonstrate that the Mp1p ELISA has the potential to detect infection earlier than culture allows and can potentially be used as a screening tool for sub-clinical infection, permitting pre-emptive antifungal therapy to prevent disease development. This is an area of active research.

Matrix-Assisted Laser Desorption/Ionization-Time of Flight (MALDI-TOF) Method

The Matrix-Assisted Laser Desorption/Ionization-Time of Flight (MALDI-TOF) method has recently been used for identification of *Talaromyces* to the species level from cultured specimens based on either an in-house database generated from institution's *T. marneffeii* clinical strain collection^{43,44} or from the comprehensive National Institutes of Health MDL Mold Library.⁴⁵ The MALDI-TOF represents a rapid and reliable tool for downstream fungal identification, eliminating the need to demonstrate thermal dimorphism.

Antifungal Susceptibility Testing

The minimum inhibitory concentrations (MIC) have been consistently low for itraconazole, intermediate for amphotericin B, and high for fluconazole. Thus far, only one retrospective case series from Chiang Mai in Thailand correlated MIC data of 30 clinical isolates with patient outcomes. More recent studies reported

low MIC values for the newer generation azole drugs voriconazole (MICs 0.016 µg/mL–0.063 µg/mL) and posaconazole (MICs 0.001 µg/ml–0.002 µg/ml), and intermediate to high MIC values of 2 µg/ml to 8 µg/mL for anidulafungin,⁴⁶ the later study utilized a commercial Sensititre YeastOne™ YO10 assay.⁴⁷ These results suggest promising activity of voriconazole and posaconazole for the treatment of talaromycosis, and suggest that the echinocandins are less effective against *T. marneffeii*.

Preventing Exposure

Two case-controls studies in Thailand and Vietnam demonstrated that patients with World Health Organization (WHO) Stage 4 HIV disease or a CD4 count <100 cells/mm³ who had an occupational exposure to plants and farmed animals were at increased risk for infection.^{20,21} The risk was higher in the rainy and humid months.^{3,22}

Residency or a history of traveling to the highland regions (as short as 3 days) was a risk factor for talaromycosis in patients with advanced HIV disease in southern Vietnam.²⁰ These data suggest that patients with advanced HIV should avoid visiting the areas where talaromycosis is highly endemic, particularly highland regions during the rainy and humid months (**BIII**).

Preventing Disease

Primary prophylaxis has been shown to reduce the incidence of talaromycosis and other invasive fungal infections. A double-blind, placebo-controlled trial in Chiang Mai, Thailand, demonstrated that oral itraconazole 200 mg daily for primary prophylaxis significantly reduced the occurrence of invasive fungal infections (predominantly cryptococcosis and talaromycosis) in patients with HIV with a CD4 count <200 cells/mm³.⁴⁸

In a retrospective study also in Chiang Mai, fluconazole (400 mg weekly) was shown to be as effective as itraconazole (200 mg daily) for primary prophylaxis.⁴⁹ However, these studies were conducted prior to the widespread use of ART, had small sample sizes, and a mortality benefit was not observed.

Therefore, primary prophylaxis has not been widely adopted given concerns about long-term toxicity, drug-drug interactions, and costs.

Indication for Primary Prophylaxis

Primary prophylaxis is only recommended for patients with HIV with CD4 counts <100 cells/mm³ who reside in the highly-endemic regions in northern Thailand, southern China, and northern and southern Vietnam who are unable to have ART for whatever reasons or have treatment failure without access to effective antiretroviral options (**BI**). The drug choices for prophylaxis are oral itraconazole 200 mg once daily (**BI**) or oral fluconazole 400 mg once weekly (**BII**).

Primary prophylaxis is not recommended in patients who are on or about to start effective ART and is not recommended in geographic areas outside of the mentioned highly endemic regions (**AIII**).

For patients with HIV from the United States and from countries outside of the endemic region who are not on effective ART, have a CD4 count <100 cells/mm³, and must travel to the highly-endemic areas mentioned, primary prophylaxis with either itraconazole or fluconazole should begin 3 days prior to travel to allow serum drug level to reach steady state and may continue for 1 week after travel (**BIII**).

Discontinuation of Primary Prophylaxis

Primary prophylaxis for talaromycosis can reasonably be discontinued in patients who are ART adherent and have a sustained CD4 count ≥100 cells/mm³ for over 6 months (**BII**). In areas where viral load monitoring has replaced CD4 count monitoring, primary prophylaxis can reasonably be discontinued in patients who achieve sustained virologic suppression over 6 months (**BIII**).

Treating Disease

Disseminated talaromycosis is fatal if untreated.⁵⁰

The case fatality rates with antifungal therapy range from 10% to 30%.^{2,3,6,16}

Antifungal therapy for talaromycosis is divided into induction, consolidation, and maintenance phases. The treatment recommendations are based on several observational studies in Thailand and China⁵¹⁻⁵⁴ and the recent Itraconazole versus Amphotericin B for Penicilliosis (IVAP) randomized, controlled trial in Vietnam.⁵⁵

In an earlier non-comparative prospective study of 74 patients in Thailand, induction therapy with deoxycholate amphotericin B for 2 weeks, followed by consolidation therapy with itraconazole for 10 weeks was shown to be highly effective. Treatment success rate (defined by negative blood culture and resolution of fever and skin lesions at the end of a 12-week treatment course) was 97%.⁵¹

Voriconazole has been used for induction therapy in patients who could not tolerate amphotericin B and was shown to have favorable clinical and microbiological outcomes in 8 of 9 patients in Thailand⁵³ and 10 of 14 patients in China.⁵²

The IVAP trial randomized 440 patients across 5 hospitals in Vietnam and demonstrated that induction therapy with amphotericin B was superior to itraconazole with respect to 6-month mortality (absolute risk of death was 11% and 21%, respectively; hazard ratio of death in the itraconazole arm was 1.88 [95% confidence interval, 1.15—3.09, $P = 0.012$]). Patients in the amphotericin B arm had significantly lower rates of disease complications, including disease relapse and immune reconstitution inflammatory syndrome (IRIS), and had a four-fold faster rate of blood fungal clearance. The difference in mortality between the arms was not dependent on disease severity (based on positive blood culture, blood fungal count, or requirement for oxygen support at presentation) or by a participant's immune status (CD4 count <50 cells/mm³ or ≥ 50 cells/mm³), ART status, or intravenous (IV) drug use.⁵⁵

The recommended induction therapy for all patients, regardless of disease severity, is amphotericin B, preferably liposomal amphotericin B 3 to 5 mg/kg body weight/day where available, or deoxycholate amphotericin B 0.7 mg/kg body weight/day, IV for 2 weeks (**AI**).

Induction therapy should be followed by consolidation therapy with oral itraconazole, 200 mg every 12 hours for a subsequent duration of 10 weeks (**AI**).⁵⁵ After this period, maintenance therapy (or secondary prophylaxis) with oral itraconazole 200 mg/day is recommended to prevent recurrence until the CD4 count rises above 100 cells/mm³ for ≥ 6 months (**AI**).⁵⁶

For patients unable to tolerate any form of amphotericin, induction therapy with IV voriconazole 6 mg/kg every 12 hours on day 1 (loading dose), then 4 mg/kg every 12 hours or with oral voriconazole 600 mg every 12 hours on day 1 (loading dose), then 400 mg every 12 hours for 2 weeks is recommended (**BII**).^{52,53}

Thereafter, either oral voriconazole or oral itraconazole 200 mg every 12 hours can be used for consolidation therapy for 10 weeks, followed by itraconazole 200 mg/day for secondary prophylaxis. The optimal dose of voriconazole for secondary prophylaxis beyond 12 weeks has not been studied.

Itraconazole is not recommended as an induction therapy for talaromycosis, regardless of disease severity (**AI**).⁵⁵

Special Considerations with Regard to Starting ART

No studies exist regarding the optimal time to start ART in patients with HIV who have talaromycosis. In the IVAP trial, the median time to ART initiation, which was similar in both arms, was 3 weeks (range: 1 week–5 weeks).

Paradoxical IRIS events occurred only in the itraconazole arm (in 11.4% of patients), suggesting

that ART can be safely initiated as early as 1 week after starting effective antifungal therapy with amphotericin B (**BIII**).⁵⁵

Monitoring of Response to Therapy and Adverse Events (Including IRIS)

Adverse Event Monitoring

Patients treated with amphotericin B should be monitored for infusion-related adverse reactions (fever, rigors, nausea, vomiting), electrolyte disturbances (particularly hypokalemia and hypomagnesemia), nephrotoxicity (rise in creatinine), and anemia. Hydration with 500 mL to 1,000 mL of normal saline and potassium supplementation before each amphotericin B infusion reduces the risk of nephrotoxicity during treatment (**AII**). Infusion-related adverse reactions can be ameliorated by pretreatment with acetaminophen and diphenhydramine.

Drug-Drug Interactions and Therapeutic Drug Monitoring

Itraconazole and voriconazole and antiretroviral (ARV) drugs such as protease inhibitors (PIs), some integrase strand transfer inhibitors (INSTIs) and non-nucleoside reverse transcriptase inhibitors (NNRTIs) can have bi-directional interactions with each other, leading to increased or decreased drug concentrations (see [Drug-Drug Interactions](#) in the [Adult and Adolescent Antiretroviral Guidelines](#)). Close monitoring is recommended when using these drugs together.

In settings where therapeutic drug monitoring (TDM) is available, serum itraconazole and voriconazole levels should be obtained in all patients to ensure adequate drug exposure (**BIII**). This is because itraconazole and voriconazole can interact with some ARV drugs, because absorption of itraconazole can be erratic, and because of the extensive interindividual variability and non-linear pharmacokinetics of voriconazole. The target serum trough concentration should be >0.5 $\mu\text{g/mL}$ for itraconazole and >1 $\mu\text{g/mL}$ for voriconazole (**BIII**). Because it is more bioavailable, itraconazole solution is preferred over the capsule formulation.

Prevention and Management of IRIS

Both unmasking and paradoxical IRIS have been described in patients with talaromycosis when ART is initiated.⁵⁷⁻⁵⁹ In the IVAP trial, 188 of 432 (44%) patients had started ART a median of 3–4 months before developing talaromycosis, indicating the role of ART in the unmasking of subclinical infection in a significant proportion of patients.⁵⁵ This finding highlights the need for a sensitive assay to screen for subclinical infection and the importance of pre-emptive antifungal therapy to prevent disease and unmasking IRIS. In patients starting ART after a diagnosis of talaromycosis, paradoxical IRIS events only occurred in patients treated with itraconazole induction therapy,⁵⁵ demonstrating the role of effective induction therapy with amphotericin B in the prevention of paradoxical IRIS. ART should not be withheld because of concerns for possible development of IRIS (**AIII**).

Patients with paradoxical IRIS typically present with inflammatory manifestations that include erythematous or immunological skin lesions such as erythema nodosum, large and painful peripheral lymph nodes, and synovitis of small joints. Most symptoms can be managed by judicious use of non-steroid anti-inflammatory medicine. Corticosteroids are reserved for synovitis that interferes with daily function.⁵⁹ Although the IRIS events in the IVAP trial were not associated with increased mortality and were managed effectively with continuation of ART and antifungal therapy, they were associated with higher morbidity, including lower quality of life and increased diagnostic testing, duration of hospitalization, and cost.⁵⁵

Managing Treatment Failure and Relapse

Talaromycosis treatment failure and disease relapse were associated with ineffective induction therapy with itraconazole, highlighting the importance of amphotericin B induction therapy.⁵⁵ On the basis of

case series that included very few patients and on clinical experiences, voriconazole is an alternative therapy for patients who are unable to tolerate amphotericin B treatment (**BII**).

Disease relapse is associated with higher mortality⁵⁵ and occurs mainly in patients who are not adherent to ART or have virologic failure, as well as in those who are not adherent to itraconazole consolidation or maintenance therapy. Therapy adherence counseling and TDM for itraconazole and voriconazole, if available, are recommended (**AIII**).

Preventing Recurrence

When to Start Secondary Prophylaxis/Chronic Maintenance Therapy

A study showed that >50% of patients not treated with ART had disease relapse within 6 months after discontinuation of antifungal therapy. A double-blind, placebo-controlled study conducted in Chiang Mai, Thailand, demonstrated that secondary prophylaxis with oral itraconazole 200 mg daily in patients with AIDS reduced the talaromycosis relapse rate from 57% to 0% ($P < 0.001$).⁵⁶ All patients who successfully complete induction and consolidation treatment for talaromycosis should receive secondary prophylaxis (maintenance therapy) with oral itraconazole 200 mg/day until they reach criteria for stopping secondary prophylaxis (**AI**).

When to Stop Secondary Prophylaxis/Chronic Maintenance Therapy

No randomized, controlled study has demonstrated the safety of discontinuation of secondary prophylaxis for talaromycosis. However, a retrospective cohort study reported no relapse of talaromycosis after itraconazole was discontinued in patients receiving ART whose CD4 counts were >100 cells/mm³.⁶⁰

Therefore, secondary prophylaxis for talaromycosis can be discontinued in patients who are ART adherent and have CD4 counts >100 cells/mm³ for at least 6 months (**BII**).

Secondary prophylaxis can reasonably be discontinued in patients with sustained virologic suppression for ≥ 6 months (**BIII**).

Secondary prophylaxis/chronic maintenance therapy should be reintroduced if the CD4 count decreases to <100 cells/mm³ (**BIII**).

Special Considerations During Pregnancy

The diagnosis and treatment of talaromycosis during pregnancy is similar to that in non-pregnant adults, with the following considerations regarding antifungal use in pregnancy. Amphotericin B has not been shown to be teratogenic in animals, and no increase in fetal anomalies has been seen with its use in humans. Neonates born to women on chronic amphotericin B at delivery should be evaluated for renal dysfunction and hypokalemia.

Itraconazole at high doses has been shown to be teratogenic in animals, but because humans lack the metabolic mechanism accounting for these defects, the animal teratogenicity data are not applicable to humans. Case series in humans do not suggest an increased risk of birth defects with itraconazole, but experience is very limited.⁶¹

Voriconazole is Food and Drug Administration Category D because of teratogenicity (cleft palate and renal defects) seen in rats and embryotoxicity in rabbits. No human data on use of voriconazole are available, so use in the first trimester is not recommended.

Substitution of amphotericin B for high-dose azoles in the first trimester is recommended (**BIII**). Women on secondary prophylaxis with itraconazole or other azoles should postpone pregnancy until their CD4 counts have been restored with ART, such that prophylaxis can be discontinued (**BIII**). If women become pregnant while receiving itraconazole prophylaxis, the decision as to whether to continue should be individualized based on current CD4 count and viral suppression and patient preference.

Recommendations for Preventing and Treating Talaromycosis

Preventing First Episode of Talaromycosis (Primary Prophylaxis)

Indication for Primary Prophylaxis:

- Persons with a CD4 count <100 cells/mm³, who are unable to have ART, or have treatment failure without access to effective ART options and who either:
 - Reside in the highly endemic regions in northern Thailand, throughout Vietnam, and southern China (particularly in highland regions during the rainy humid months) **(BI)**, or
 - Are from countries outside of the endemic region and must travel to the region **(BIII)**.

Primary Prophylaxis

For Individuals Residing in Endemic Areas:

- Preferred Therapy: Itraconazole 200 mg PO once daily **(BI)**
- Alternative Therapy: Fluconazole 400 mg PO once weekly **(BII)**

For Individuals Traveling to Endemic Areas:

- Preferred Therapy: Begin itraconazole 200 mg PO once daily 3 days before travel and continue for 1 week after leaving the endemic area **(BIII)**.
- Alternative Therapy: Begin fluconazole 400 mg 3 days before travel, then continue 400 mg once weekly while in the area, and take final dose after leaving the endemic area **(BIII)**.

Indication for Discontinuing Primary Prophylaxis for Persons who Reside in Endemic Areas:

- CD4 count >100 cells/mm³ for ≥ 6 months in response to ART **(BII)**
- Viral load suppression for ≥ 6 months on ART **(BIII)**

Indication for Restarting Primary Prophylaxis:

- CD4 count decreases to <100 cells/mm³ **(BIII)** and patient still resides in or travels to high-risk areas. Primary prophylaxis for travelers may begin three days prior to travel to allow serum drug level to reach steady state and may continue for one week after travel **(BIII)**.

Treating Acute Infection in Severely Ill Patients

Preferred Therapy:

- Induction therapy with liposomal amphotericin B 3 to 5 mg/kg/day IV for 2 weeks, followed by consolidation therapy with itraconazole 200 mg PO twice daily for 10 weeks **(AI)**, followed by maintenance therapy or secondary prophylaxis with itraconazole 200 mg PO daily **(AII)**

Alternative Therapy:

- In settings where liposomal amphotericin B is not available, induction therapy with deoxycholate amphotericin B 0.7 mg/kg/day IV for 2 weeks, followed by consolidation therapy with itraconazole 200 mg PO twice daily for 10 weeks **(AI)**, followed by maintenance therapy or secondary prophylaxis with itraconazole 200 mg PO daily **(AII)**
- In settings where amphotericin B is not available, induction therapy with voriconazole 6 mg/kg IV every 12 hours for 1 day (loading dose) and then voriconazole 4 mg/kg IV every 12 hours for 2 weeks, or oral voriconazole 600 mg every 12 hours on day 1 (loading dose) and then voriconazole 400 mg PO every 12 hours for 2 weeks; followed by consolidation therapy with voriconazole 200 mg PO twice daily or itraconazole 200 mg PO twice daily for a maximum of 10 weeks **(BII)**; followed by maintenance therapy or secondary prophylaxis with itraconazole 200 mg PO daily **(AII)**
- Itraconazole is not recommended as induction therapy for talaromycosis **(AI)**.

Criteria for Discontinuing Chronic Maintenance Therapy:

- CD4 count >100 cells/mm³ for ≥ 6 months in response to ART **(BII)**
- Virologic suppression for ≥ 6 months on ART **(BIII)**

Criteria for Restarting Chronic Maintenance Therapy:

- CD4 count decreases to <100 cells/mm³ **(AIII)**

Other Considerations

- ART can be initiated as early as one week after the initiation of treatment for talaromycosis with amphotericin B induction therapy to improve outcomes **(BII)**.
- Given erratic absorption of itraconazole, extensive interindividual variability and non-linear PKs of voriconazole, and the potential for drug interactions with ARV drugs, itraconazole and voriconazole concentrations should be monitored, and serum trough concentration should be >0.5 $\mu\text{g/mL}$ for itraconazole and >1 $\mu\text{g/mL}$ for voriconazole **(BIII)**. Both itraconazole and voriconazole can have significant drug-drug interactions with various ARV drugs; dosage adjustment may be necessary, and TDM to guide therapy can be considered (see the [Drug-Drug Interactions](#) tables in the [Adult and Adolescent Antiretroviral Guidelines](#) for further recommendations).

Key: ART = antiretroviral therapy; ARV = antiretroviral; CD4 = CD4 T lymphocyte; IV = intravenous; PK = pharmacokinetic; PO = orally; TDM = therapeutic drug monitoring

Figure 1. Geographic Distribution of Talaromyces

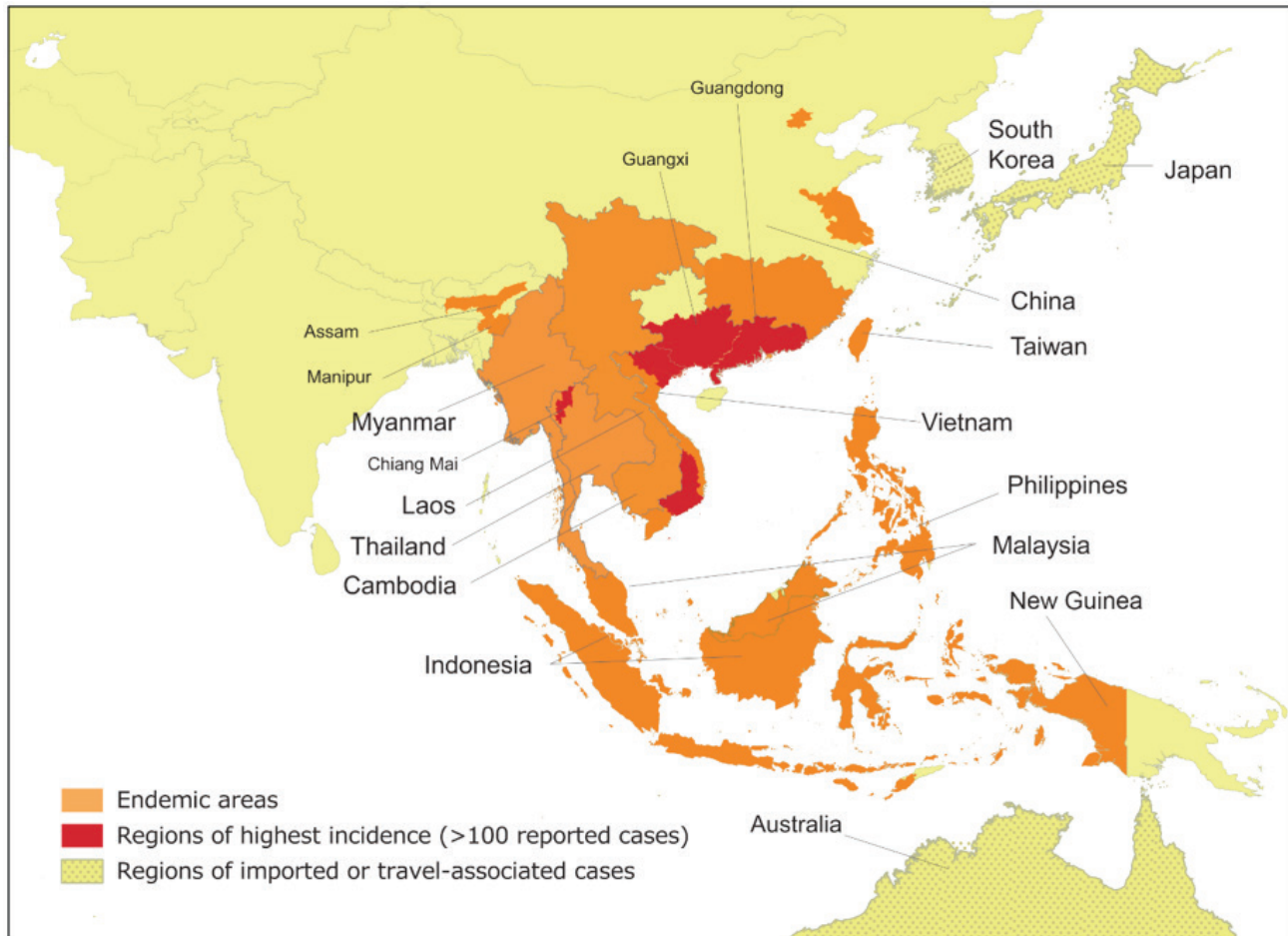


Figure courtesy of Dr. Thuy Le, Division of Infectious Diseases and International Health, Duke University School of Medicine

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